A Novel Mouse Model of Inflammatory Bowel Disease Links Mammalian Target of Rapamycin-Dependent Hyperproliferation of Colonic Epithelium to Inflammation-Associated Tumorigenesis

Inflammatory bowel disease (IBD) is a high-risk condition for human colorectal cancer. However, our mechanistic understanding of the link between inflammation and tumorigenesis in the colon is limited. Here we established a novel mouse model of colitis-associated cancer by genetically inactivating signal transducer and activator of transcription 3 (Stat3) in macrophages, with partial deletion in other myeloid and lymphoid cells. Inflammation developed in the colon of mutant mice spontaneously, and tumor lesions, including invasive carcinoma, arose in the inflamed region of the intestine with a frequency similar to that observed in human IBD patients. The development of both inflammation and tumors in the mutant mice required the presence of microflora. Indeed, inflammation was associated with disruption of colonic homeostasis, fulminant epithelial/tumor cell proliferation, and activation of the mammalian target of rapamycin (mTOR)-Stat3 pathway in epithelial and tumor cells. The activation of this pathway was essential for both the excess proliferation of epithelial/tumor cells and the disruption of colonic homeostasis in the mutant mice. Notably, a similar abnormal up-regulation of mTOR-Stat3 signaling was consistently observed in the colonic epithelial cells of human IBD patients with active disease. These studies demonstrate a novel mouse model of IBD-colorectal cancer progression in which disrupted immune regulation, mTOR-Stat3 signaling, and epithelial hyperproliferation are integrated and simultaneously linked to the development of malignancy.

Chronic inflammation in the intestine ranks among the top three high-risk conditions for colorectal cancer. In contrast to the other high-risk conditions, the hereditary syndromes of familial adenomatous polyposis and hereditary nonpolyposis colorectal cancer, in which the genetic predispositions have been identified, the mechanistic link between inflammation and tumorigenesis is still largely unknown. IBD is thought to be a unique modulation of the intestinal mucosal immune response determined by a complex interplay of genetic, microbial, and environmental factors. Animal models that mimic human IBD and its associated colorectal cancers have contributed to recent progress in the understanding of mucosal immunity and its dysregulation in tumorigenesis. A number of widely used models involve chemically induced inflammation-associated colorectal cancers, initiated by injecting mice

Supported by a Montefiore Medical Center New Research Initiative Award, a Miriam Mandel Faculty Scholar Award, the National Cancer Institute (NCI) (R01-94173, P01-CA100324, and U54-CA100926), and Albert Einstein Cancer Center Core grant P30-CA13330. The content is solely the responsibility of the authors and does not necessarily represent the official views of the NCI.

A guest editor acted as editor-in-chief for this manuscript. No person at Thomas Jefferson University or Albert Einstein College of Medicine was involved in the peer review process or final disposition for this article.

Accepted for publication October 1, 2009.

Supplemental material for this article can be found on http://ajp.amjpathol.org.

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with carcinogens such as azoxymethane and/or introducing chemical irritants such as dextran sulfate sodium. Such treatments lead to acute inflammation in the intestine promoted by mucosal damage induced by the chemical, rather than chronic inflammation as observed in patients with IBD, and therefore may not recapitulate stochastic events involved in inflammation-associated cancers in the human.

An alternative approach is to model IBD by genetic modifications that favor the development of chronic intestinal inflammation. Signaling through interleukin (IL)-10, an anti-inflammatory cytokine, has been targeted, because evidence indicates that this cytokine is important in maintaining the hyperresponsive state of the intestinal mucosa to normal bacterial microflora. Sequence variants or polymorphisms in the IL-10 gene have been reported to contribute to IBD susceptibility, indicating that interference with IL-10 regulation may make an important contribution to the etiology of IBD. Indeed, inflammation and tumors develop in the colon of IL-10+/− mice in a manner dependent on the presence of intestinal microflora. However, because IL-10 deletion is present in all cells in these mice, it is difficult in this model to distinguish effects due to dysregulation of stromal and immune cells from effects arising from IL-10 mutation in the epithelium.

To address this issue, attempts have been made to inactivate IL-10 signaling more specifically in cells of the immune system. Signal transducer and activator of transcription 3 (Stat3) plays a major role in Janus tyrosine kinase/Stat signaling induced by IL-10. Mice with inactivation of Stat3 targeted by interferon-responsive Mx1-Cre to multiple types of cells including macrophages and gut epithelial cells or targeted to myeloid cells with LysM-Cre, developed colitis in the intestine. However, unlike in IL-10+/− mice, targeting of Stat3 inactivation in these mice was not reported to cause tumor formation, suggesting that additional factor(s) are required for prompting tumor development in the colon in the setting of an inflammatory response. In this regard, although IL-10 plays a major role in myeloid cells, the cytokine also inhibits activation and function of T cells, granulocytes, B cells, and natural killer cells, although mice with T lymphocyte-specific depletion of Stat3 did not develop either colitis or colonic tumors.

We therefore tested the effect of the conditional inactivation of Stat3+ genes in immune cells using Cre driven by the colony-stimulating factor-1 receptor promoter. This inactivated Stat3 in both myeloid and lymphoid cells, including macrophages, lymphocytes, and granulocytes, which produced not only a dramatic inflammatory response of the intestine but also eventual malignant tumor formation at a frequency similar to that observed in human IBD patients. These phenotypes were completely dependent on the intestinal microflora. This novel mouse model of human IBD recapitulates the increased probability of tumor development in the colon, an important clinical phenotype in human IBD. Furthermore, using this model we present the first evidence that the inflammation-associated hyperproliferation of colonic epithelial cells is dependent on the activation of signaling through an mTOR-Stat3 pathway.

**Materials and Methods**

**Mice**

All procedures were conducted in accordance with National Institutes of Health regulations concerning use and care of experimental animals and approved by the Albert Einstein College of Medicine animal use committee. All of the mice were housed in the pathogen-free barrier facility at Albert Einstein College of Medicine.

**Stat3-IKO Mice**

To establish the transgenic construct for Tg(Csf1r-iCre)jwp, the improved Cre (iCre) sequence was inserted into Apal and NotI sites using blunt-ended ligation downstream from the colony-stimulating factor-1 receptor promoter. The transgenic mice were generated using standard methods in the Transgenic Mouse Facility at Albert Einstein College of Medicine. Tg(Csf1r-iCre)jwp mice were identified by PCR-based genotyping with the following primers: F1, 5’-CAGGCGCTTCCACACCAGC-3’; and F2, 5’-CTGGCTGTGAAGACCATC-3’. Stat3flox/flox mice (C57BL/6) were kindly provided by Dr. Colin Stewart (National Institutes of Health), and mice were identified using a standard PCR-based genotyping procedure with following primers: F1, 5’-GCTGCCAACAGCCACTGC-CCCAG-3’; F2, 5’-GAAGGCAGGTCTCTCTGGTGCTTC-3’; and F3, 5’-CAGAACCGGCGGCTCAGTGC-3’. The PCR reaction yields a 140-bp product from wild-type, a 260-bp product from the flox, or a 379-bp product from the flox-depleted sequence of the Stat3 gene. The mice used in the experiments were at or more than three generations of backcrossing to C57BL/6 mice.

**Flow Cytometry**

Bone marrow-derived macrophages (BMMs) were prepared as described previously with minor changes. Antibodies used were R-phycocerythrin- or allophycocyanin-conjugated rat anti-mouse F4/80 (Invitrogen, Carlsbad, CA), R-phycocerythrin-conjugated rat anti-mouse CD3 (Invitrogen), R-phycocerythrin-conjugated rat anti-mouse Ly6G (BD Pharmingen, San Diego, CA), PerCP-conjugated rat anti-CD11b (BD Pharmingen), mouse anti-mouse Stat3 (IgG2a) (Cell Signaling Technology, Danvers, CT), and Alexa Fluor 488-conjugated goat anti-mouse IgG2a. For each reaction, isotopic nonspecific antibody and mouse IgG2a were used as controls for the primary antibody and secondary antibody only controls were used.

**Western Blotting Analysis**

Western blotting analysis was performed following the standard procedure. Antibodies were anti-Stat3 (1:2000,
Cell Signaling), anti-β-actin (2 ng/ml, Sigma-Aldrich, St. Louis, MO), and anti-Bcl-xL (1:1000, Cell Signaling).

**Human Colon Tissue**

Human colon sections from four individuals with active IBD were provided by the Pathology Department at Montefiore Medical Center (institutional review board exempted protocol 09-03-096X). The tissue array for human normal and sporadic colon cancer was from Imgenex (IMH-359, Imgenex, San Diego, CA).

**Histological Analysis**

Colons were prepared as “Swiss rolls,” fixed in 10% formalin, and then paraffin-embedded. Sections were stained with H&E. The degree of inflammation and tumors of the colon sections were evaluated by R.S.S., a veterinary pathologist in the Albert Einstein Cancer Center Histology Core Facility. Inflammation was scored as follows: 0, no inflammation; 1, minimal; 2, mild; 3, moderate; 4, marked; and 5, severe. Lesions were classified (evaluated in regard to the consensus statement of mouse model of intestinal cancers, with modifications\(^\text{20}\)) as follows. **Hyperplasia** was characterized by thickened mucosa with elongated crypts. Crypts consist of goblet cells and morphologically normal epithelium with nuclear morphology and polarity similar to those of the normal controls. **Dysplasia** (hyperplasia with dysplasia and inflammation) was classified by thickened mucosa with elongated, irregularly branched glands that contain epithelium with cytological and nuclear atypia including loss of differentiation (goblet cells), polarity, and enlarged nuclei. Areas of the intestine with inflammation graded as moderate or greater generally had some degree of dysplasia. Dysplasia was graded in five levels similar to the inflammation score described above. **Carcinoma** is distinguished by tumor cell invasion into submucosa. Specific features also include the presence of irregular glands, desmoplasia, and cell loss as well as multiple invading sites in the invading mucosa.\(^\text{20}\) The leukocytic infiltration and injury scores were determined following the procedure of Xiao el al.\(^\text{21}\) For goblet cell identification, tissue sections were stained with 1% Alcian blue (pH 2.5) and 0.1% neutral red following standard procedures. For quantitative histological analysis, the stained tissue sections were photographed to produce TIFF images using an Olympus IX70 microscope and a Sesicam QEcooled charge-coupled device camera. The entire inflamed colon was photographed, and at least three mice per group were analyzed. Positively stained areas were measured and normalized with the areas of nuclei in the same fields using ImageJ software as described previously.\(^\text{22}\) Mucosal thickness was measured as the length from the bottom of the crypt to the tip of the gland in TIFF images using ImageJ. Complete glands of colon sections (approximately seven per mouse) were measured in the inflamed region of Stat3-IKO mice and the corresponding region in Stat3\(^{\text{flox/flox}}\) mice.

**Yellow Fluorescence Protein Detection**

Tissue sections from Tg(Csf1r-iCre)jwp.Rosa-YFP mice were prepared as described previously.\(^\text{22}\) Sections were counterstained with 4',6'-diamidino-2-phenylindole and imaged using a Leica SP2 AOBS laser scanning confocal microscope in the Analytical Image Facility at Albert Einstein College of Medicine. The yellow color of yellow fluorescence protein (YFP) was converted to green using Adobe Photoshop.

**Immunohistochemistry**

Tissues for immunohistochemistry (IHC) experiments were fixed with 10% formalin, except for those for F4/80 IHC, which were fixed in Zinc fixative (BD Pharmingen), using standard procedures. Antibodies were 0.1 μg/ml anti-F4/80 (Invitrogen), 3 μg/ml anti-CD3 (DakoCytomation, Carpinteria, CA), 5 μg/ml anti-Ly-6-G, 1:200 of anti-Ki-67, 4 μg/ml anti-pStat3 (Tyr705), 1:100 for anti-cyclin D1 (Lab Vision, Fremont, CA), 78 ng/ml anti-CD31 (BD Pharmingen), 1:50 for anti-pStat3 (Ser-727), 1:400 for anti-pS6, and 1:400 for anti-Survivin (Cell Signaling). The standard IHC procedure provided by Cell Signaling was used.

**Antibiotic Treatment**

The published procedure for treatment of mice with a combination of antibiotics (vancomycin, neomycin, metronidazole, and ampicillin [VNMA]) and the detection of bacteria in stool was used.\(^\text{21}\) Mice were treated with 10 mg/kg daily Everolimus (Sigma-Aldrich) or 1% dimethyl sulfoxide/saline for 3 days by gavages.

**Results**

**Stat3 Conditionally Depleted Mice**

To determine the effect of blocking Stat3-mediated signaling in macrophages, we established a transgenic mouse line, Tg(Csf1r-iCre)jwp, in which expression of Cre was under control of the colony-stimulating factor-1 receptor promoter (Csf1r or Fms).\(^\text{15,16}\) An improved Cre (iCre) sequence, based on mammalian codon usage,\(^\text{17}\) was used for the transgenic construct to maximize efficiency of target excision. After introduction into FVB embryos, one of three founder mice transmitted the transgene to its offspring, which were phenotypically normal. The Tg(Csf1r-iCre)jwp transgene was then crossed into Rosa-26-YFP reporter mice to examine iCre-dependent deletion at this marker locus. Analysis of Tg(Csf1r-iCre)jwp reporter mice as an iCre-dependent deletion at this marker locus. Analysis of Tg(Csf1r-iCre)jwp reporter mice revealed that the YFP signal was detected only in mice that carried the transgene. Signal was seen in various tissues, including liver, spleen, intestine, heart, kidney, and muscle; in each tissue, expression was found in cells with characteristics consistent with macrophages/dendritic cells (supplemental Figure S1A, see http://ajp.amjpathol.org). Consistent with these data, fluorescence-activated cell sorter
analysis revealed YFP signal in ~80% of BMMs (supplemental Figure S1Ba, see http://ajp.amjpathol.org). However, a YFP signal was also found in ~50% of bone marrow granulocytes and splenic T lymphocytes (Supplemental Figure S1Bb and c, see http://ajp.amjpathol.org). The transgene was then crossed into mice that were Stat3flox/flox, targeting excision of exons 16 to 21 of the Stat3 gene, thus inactivating the protein. Consistent with the above data, efficient deletion in Stat3 was observed in BMMs in Tg(Csf1r-iCre)jwp mice (Supplemental Figure S1C, see http://ajp.amjpathol.org), reducing the level of Stat3 protein in BMMs of these mice to ~20% of that present in BMMs from Tg(Csf1r-iCre)jwp/mice (Figure 1A). Fluorescence-activated cell sorter analysis confirmed efficient Stat3 depletion in BMMs and relatively lower efficiency (~50%) in granulocytes and T or B lymphocytes isolated from Tg(Csf1r-iCre)jwp/mice compared with those from Stat3flox/flox mice (data not shown). Because Stat3 depletion was found in inflammatory cells rather than in the intestinal epithelium (described below), Tg(Csf1r-iCre)jwp mice were referred to as Stat3-IKO. Because no Stat3 depletion was found in Tg(Csf1r-iCre)jwp mice that did not carry the transgene and they were phenotypically normal, these were subsequently used as the wild-type control (ie, Stat3ff controls).

Development of Spontaneous Colitis in Stat3-IKO Mice

Stat3-IKO mice (on a mixed background of FVB/C57BL/6 backcrossed for at least three generations to Stat3flox/flox mice) were obtained at the expected Mendelian ratio with no abnormalities at birth or within the first few weeks after parturition (Figure 1A), indicating that
inactivation of Stat3 by the transgene did not detectably alter embryonic or early development. However, ~20% of Stat3-IKO mice developed pronounced colitis at 8 weeks of age, which increased to ~50% by 18 to 20 weeks of age. These mice exhibited persistent diarrhea, bloody feces, and weight loss. The remaining mutant mice, although without these prominent symptoms, also were less physically active and had significantly lower body weights at older ages compared with age-matched Stat3f/f controls (Figure 1, B and E). Histological analysis of Stat3-IKO with or without evident colitis revealed increased leukocytic infiltration and intestinal mucosal thickening compared with Stat3f/f littermates (Figure 1, C versus D), predominantly in the proximal colon and cecum, without gross pathological abnormalities in the small intestine (data not shown). Consistent with these observations, the leukocytic infiltration score (degree of leukocytic infiltration and increased mucosal thickness)23 was significantly higher in the Stat3-IKO colons compared with control colons (Figure 1F). Twenty of 21 Stat3-IKO mice aged 8 to 39 weeks developed inflammation in the colon, whereas only 1 of 21 Stat3f/f controls exhibited a mild increase in leukocytic infiltration (Table 1, Fisher’s exact test, P < 0.0001).

Macrophages (F4/80+ cells) markedly increased in the colon of Stat3-IKO mice (Figure 1, I and K versus J and L). There was also a striking increase in CD3+ T-cells (Figure 1, M and O versus N and P) and a moderate increase in granulocytes (Figure 1, Q versus R). Quantitative analysis by IHC showed a significantly higher density of F4/80+ and CD3+ cells in the proximal colon of Stat3-IKO mice (Figure 1, G and H, respectively). Thus, inactivation of Stat3 in Stat3-IKO mice was associated with an inflammatory reaction specifically in the colon.

Tumor Development in Inflamed Colon and Cecum in Stat3-IKO Mice

Visible polyps were found in the inflamed areas of the proximal colon (Figure 2A, arrows) and cecum (data not shown). As with tumors found in human IBD,23 most of the lesions in Stat3-IKO mice were “flat” with a broad base (Figure 2C) or morphologically similar to “dysplasia-associated lesions or masses.”24 Carcinoma was identified in some of these nodules, with tumor invasion to submucosa or even muscularis externa (Figure 2D, arrow). In addition to its invasion into the submucosa or deeper, carcinoma was identified by specific features described in the consensus report and recommendations for pathology of mouse models of intestinal cancer.20 These included the presence of irregular or angulated glands with cell loss in invasive components (Figure 2E, arrows), desmoplasia (Figure 2D and Supplemental Figure S2a and c, see http://ajp.amjpathol.org) and multiple invading mucosa (Supplemental Figure S2c, see http://ajp.amjpathol.org). Further, cells in invasive glands exhibited severe atypia with discernible pleomorphism and loss of the basement membrane (Supplemental Figure S2b and d, see http://ajp.amjpathol.org). These features are distinct from hernias of colonic mucosa also found in the inflamed colon and cecum in Stat3-IKO mice (data not shown). In addition to carcinoma, hyperplasia (Figure 2, H–I) and dysplasia (Figure 2, J and K) were identified in the inflamed intestine in Stat3-IKO mice as described in detail in Materials and Methods. Neoplastic lesions were found to be restricted to inflamed areas of the colon or cecum, with 63% of 8- to 39-week-old Stat3-IKO mice exhibiting dysplasia and 16% of Stat3-IKO mice developing carcinoma (Table 1). None of the age-matched Stat3f/f controls developed dysplasia or carcinoma, a highly significant difference between the groups (Fisher’s exact test, P < 0.0001). The dysplastic lesions were further graded 1 to 5 in regard to tumor progression, as described in Materials and Methods. There was a significant positive correlation between the grade of dysplasia and the degree of inflammation in the colon of Stat3-IKO mice (Figure 2L), demonstrating that colon tumor development in Stat3-IKO mice was closely associated with the presence of inflammation.

Inflammation and Tumor Development in Stat3-IKO Colon Are Dependent on the Intestinal Microflora

Disrupted interaction of the host immune system with the commensal microflora, a major component of the intestinal microenvironment that contributes to mucosal homeostasis,25 probably contributes to the development of human IBD.26 To determine whether inflammation and tumor development in Stat3-IKO mice were dependent on the intestinal microflora, Stat3-IKO and Stat3f/f control mice were treated with a combination of antibiotics (VNMA) to substantially reduce the bacterial content of the intestinal lumen.21 After 4.5 weeks of continuous administration of antibiotics through the drinking water, bacteria was found in the stool of Stat3-IKO mice than in that of the Stat3f/f controls, indicating that abnormal regulation of mucosal immunity may induce changes in microflora in colon (Figure 3A versus B). Parallel to this reduction of bacteria, both inflammation and tumor de-

| Table 1. Comparison of Tumor Development in Stat3f/f and Stat3-IKO Mice |
|-----------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Mice (8–39 Weeks)           | Inflammation    | No lesion       | Hyperplasia     | Dysplasia       | Carcinoma       |
| Stat3-IKO                   | 20/21           | 6/19 (32%)      | 13/19 (68%)     | 12/19 (63%)     | 3/19 (16%)      |
| Stat3f/f                    | 4/21            | 17/17 (100%)    | 0               | 0               | 0               |

Statistical analysis: Stat3f/f versus Stat3-IKO (Fisher’s exact test): inflammation, P < 0.0001; tumor development (dysplasia + carcinoma), P < 0.0001.
velopment in the colon of VNMA-treated Stat3-IKO mice were significantly inhibited (Table 2). There was a notable lack of inflammation and reduction of mucosal thickness in the colon of VNMA-treated Stat3-IKO mice (Figure 3, E versus F), and the histopathology of the colonic mucosa of these mice was similar to the untreated Stat3f/f controls (Figure 3, D versus F). The leukocytic infiltration score and the infiltration of both macrophages and T lymphocytes were also markedly reduced (Figure 3, G–K, and N). However, compared with those in untreated Stat3f/f control mice, infiltrated macrophages in the VNMA-treated Stat3-IKO mice were still significantly higher (Figure 3M, CON versus IKO), indicating that the treatment did not completely inhibit inflammation in the colon. The injury score, measured by the extent of loss of goblet cells in the crypt, was also consistently significantly higher in VNMA-treated Stat3-IKO colon compared with the Stat3f/f control colon (Figure 3L, CON versus IKO).

Parallel to the reduction of inflammation, the development of colonic tumors was significantly reduced by antibiotic treatment: no tumors were found in any of the 11 Stat3-IKO mice from 21 to 27 weeks of age treated with antibiotics, and only 2 of these treated mice displayed mild inflammation and hyperplasia at the end of the treatment. The tumor incidence was significantly lower than that in untreated Stat3-IKO mice 27 weeks of age or younger (Table 2, \( P < 0.001 \)). These data indicate that the inflammation and tumor development in Stat3-IKO colon was dependent on the intestinal microflora. Further investigation may reveal the specific species of bacteria involved. Because a large percentage of Stat3-IKO at 21 weeks of age has developed dysplasia, the result also suggests that the reduction in inflammation by antibiotic treatment may cause a regression of the early stages of dysplasia and neoplasia (Table 2). However, the low frequency of carcinoma precluded determination of whether the treatment had an effect on established carcinomas.
Impact of Inflammation on Intestinal Homeostasis

To explore potential mechanisms by which inflammation might have promoted tumorigenesis in the intestine of Stat3-IKO mice, we examined the impact of inflammation on colonic epithelial cell proliferation, apoptosis, and differentiation. As noted, there was a prominent alteration in mucosal thickness of the proximal colon from 120 μm in controls to >400 μm in inflamed regions in Stat3-IKO mice (Figure 4A).

Figure 3. Inflammation and tumor development in Stat3-IKO colon is dependent on intestinal microflora. A–C: Bacterial density in stools of an untreated Stat3f/f (A), untreated Stat3-IKO (B), or a VNMA-treated Stat3-IKO (C) mouse. D–F: The correlated histology of the colon. Scale bar = 100 μm. Immunohistochemical analysis for macrophages (F4/80) (G and H) and T lymphocytes (CD3) (I and J). Scale bar (in J for all of the images) = 100 μm. K: Comparison of leukocytic infiltration. *Unpaired t-test: IKO versus IKO-T, \( P = 0.019 \); CON versus IKO-T, \( P = 0.019 \). L: Epithelial injury score. *Unpaired t-test: IKO versus IKO-T, \( P = 0.019 \); CON versus IKO-T, \( P = 0.019 \). M: Quantitative immunohistochemical analysis for F4/80. *Unpaired t-test: IKO versus IKO-T, \( P < 0.001 \); CON versus IKO-T, \( P = 0.047 \). N: Quantitative immunohistochemical analysis for CD3. *Unpaired t-test: IKO versus IKO-T, \( P = 0.023 \); CON versus IKO-T, \( P = 0.29 \). CON, untreated Stat3f/f; IKO, untreated Stat3-IKO and IKO-T, VNMA-treated Stat3-IKO mice.

Table 2. Effect of Antibiotic Treatment

<table>
<thead>
<tr>
<th>Mice</th>
<th>Inflammation</th>
<th>No lesion</th>
<th>Hyperplasia</th>
<th>Dysplasia</th>
<th>Carcinoma</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stat3-IKO (8–27 weeks)</td>
<td>14/15 (93%)</td>
<td>5/15 (33%)</td>
<td>10/15 (67%)</td>
<td>10/15 (67%)</td>
<td>3/15 (20%)</td>
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<tr>
<td>Stat3-IKO VNMA-treated (21–27 weeks)</td>
<td>3/11 (27%)</td>
<td>9/11 (82%)</td>
<td>2/11 (18%)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Stat3f/f (8–27 weeks)</td>
<td>1/16 (6%)</td>
<td>16/16 (100%)</td>
<td>0</td>
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<td>0</td>
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</table>

Statistic analysis: Fisher’s exact test, two-tailed for inflammation: Stat3-IKO VNMA treated versus untreated, \( P < 0.001 \); Stat3 IKO VNMA treated versus Stat3f/f untreated, \( P = 0.55 \); for tumor development (dysplasia): Stat3-IKO VNMA treated versus untreated, \( P < 0.001 \); for tumor development (carcinoma): Stat3-IKO VNMA-treated versus untreated, \( P = 0.175 \).
lower half of the gland, as shown in Figure 4B. In the inflamed areas of the colon in Stat3-IKO mice, Ki-67+ cells were found along the entire length of the crypt, including the uppermost region adjacent to the lumen (Figure 4C, arrows), demonstrating a persistent proliferation of colonic epithelium responding to the inflammation. No obvious difference in the death of epithelial cells was found when the colons of Stat3-IKO mice were compared with those of Stat3f/f controls (data not shown).

Mature glands of the colon consist of several cell populations, including goblet cells, which can produce and secrete mucin and are a major component of the epithelium. Using Alcian blue/neutral red staining to visualize these cells, we found a significant decrease in goblet cell density in the inflamed colon of Stat3-IKO mice compared with equivalent regions in the Stat3f/f controls (Figure 4, D and F versus E and G). This finding was confirmed by quantitative analysis (Figure 4J). Consistent with these data, the injury score was an order of magnitude higher in Stat3-IKO mice than in controls (Figure 4K), demonstrating perturbed cell differentiation in the colon of Stat3-IKO mice. The depletion of goblet cells is interesting in light of the fact that genetic ablation of this lineage and the mucin they produce leads to tumor formation associated with an inflammatory response.

In addition, using an antibody that recognizes the endothelial cell-specific marker, CD31, we observed a markedly increased vessel density in the inflamed colon in Stat3-IKO mice (Figure 4, H versus I), suggesting increased angiogenesis in the inflamed tissue. Overall, these findings indicate a significant alteration of homeostasis of the colonic epithelium within the large intestine of the Stat3-IKO mice, including a marked increase in proliferating epithelial cells and a loss of colonic epithelial differentiation.
Role of Stat3 and mTOR Signaling in Tumorigenesis in the Inflamed Colon of Stat3-IKO Mice

In studies directed to oncogenic pathways that may promote tumorigenesis in the colon of Stat3-IKO mice, we found that both Stat3 and mTOR are highly activated in the epithelial and tumor cells in the inflamed colon. When the colon of Stat3f/f mice was examined using an antibody that recognizes the activated form of Stat3 (phosphorylated at Tyr705), colonic epithelium was negative for activated Stat3 (Figure 5A, black arrows), whereas some mononuclear stromal cells were positive (Figure 5A, red arrows). In contrast, colonic epithelial and tumor cells in Stat3-IKO mice (in which the Stat3 gene remains structurally intact) were positive for nuclear activated Stat3, with more intense staining in the enlarged nuclei of tumor cells (Figure 5B, black arrows). In addition to the epithelial cells, a high density of infiltrated mononuclear cells with activated Stat3 was also observed in the inflamed colon of Stat3-IKO mice (Supplemental Figure S3, see http://ajp.amipathol.org), suggesting that Stat3 is not inactivated in some subsets of myeloid and lymphoid cells involved in inflammation. Consistent with constitutive activation of Stat3 in colonic epithelium, we observed increased epithelial expression of Stat3 target genes, in-
cluding cyclin D1, Survivin, and BCL-xL, which are functionally important in driving cell proliferation and survival as well as neoplastic transformation.\textsuperscript{30} In the colon of control mice, cyclin D1 was expressed principally in the crypt epithelial cells (Figure 5, C and E, arrows), whereas in Stat3-IKO mice expression was elevated in the majority of epithelial cells throughout the entire hyperplastic gland in inflamed areas of the colon, as well as in dysplastic regions (Figure 5, D and F) and in carcinoma (data not shown). Another Stat3 target gene, Survivin, is a key regulator of mitosis and apoptosis and is tightly regulated in normal colon.\textsuperscript{31,32} In the colon of Stat3\textsuperscript{IKO} control mice, only a few Survivin-positive cells (~5 positive cells/gland) were found in crypts of the colonic glands (Figure 5G, arrows). In contrast, a large increase in Survivin-positive epithelial cells was evident in the inflamed colon and tumors of Stat3-IKO mice compared with controls (~50 positive cells/gland) (Figure 5H). Finally, we observed elevated expression of another Stat3-regulated apoptosis inhibitor, BCL-xL, in the mucosa of the proximal colon of Stat3-IKO mice compared with that in controls (Figure 5I). These results suggest that inflammation in Stat3-IKO mice may promote abnormal proliferation and survival of colonic epithelial cells through the activation of Stat3 and associated pathways in these cells.

The serine/threonine kinase mTOR is a key effector for many cellular pathways, and dysregulation of mTOR signaling has been found in various human cancers.\textsuperscript{33} To determine whether abnormal activation of mTOR was induced in the inflamed intestine in Stat3-IKO mice, we examined the phosphorylation of S6 ribosomal protein (Ser-235/236) (pS6), which is catalyzed by S6 kinase in a mTOR-dependent manner.\textsuperscript{34} Few pS6-positive epithelial cells were found in normal colon, whereas strong expression of pS6 was found in colonic epithelial cells in the inflamed region of Stat3-IKO colon (Figure 5, J and L versus K and M). This elevated expression of pS6 was restricted to areas with a high density of leukocytic infiltration (Figure 5K, black arrows), whereas very few pS6-positive cells were found in adjacent areas with milder infiltration (Figure 5K, red arrows).

To determine whether the activation of mTOR was functionally linked to the disruption of colonic homeostasis in the inflamed colon, Stat3-IKO and Stat3\textsuperscript{IKO} mice (9 to 12 weeks of age) were treated with 10 mg/kg/day Everolimus for 3 days. Everolimus is a derivative of rapamycin, a specific inhibitor of mTOR complex 1, the complex responsible for mTOR-dependent phosphorylation of S6.\textsuperscript{33} In Everolimus-treated Stat3-IKO littermates pS6 expression was at a level similar to that of control Stat3\textsuperscript{IKO} mice (Figure 5, L versus N), whereas strong expression of pS6 was observed as expected in regions of leukocytic infiltration in the colon of untreated Stat3-IKO mice (Figure 5O), confirming the effectiveness of the inhibitor. Correlated with the reduced expression of pS6, overproliferation of epithelial cells (measured by Ki-67 expression) was markedly reduced in the colon of Everolimus-treated Stat3-IKO mice compared with untreated Stat3-IKO littermates (Figure 5, P versus Q). Indeed, despite the prominent leukocytic infiltration that remained in the area (Figure 5P, arrows), the distribution of Ki-67\textsuperscript{+} cells in the colon of treated mice was similar to that of normal colon, and the cells were generally restricted to the lower half of the gland, as shown in Figure 4B. These results indicate that the mTOR pathway is activated in inflammation-associated colonic epithelium and implicate this activation in the promotion of epithelial cell proliferation.

To confirm the role of inflammation in the activation of Stat3 and mTOR, we assessed activated Stat3 and pS6 in Stat3-IKO mice treated with VNMA to remove intestinal microflora. A marked decrease in both Stat3 activation and pS6 expression was observed in the colonic epithelial cells of VNMA-treated compared with untreated Stat3-IKO mice (data not shown).

Regulation of Stat3 Activation in Epithelial Cells in the Inflamed Colon by mTOR

We next asked whether there was a functional link between mTOR and Stat3 activation in the inflamed colon. We observed that Everolimus treatment of Stat3-IKO mice led to a marked reduction in the activation of Stat3 in colonic epithelial cells, despite the prominent leukocytic infiltration that remained in the area (Figure 6A, arrows). As expected, untreated Stat3-IKO littermates displayed activated Stat3 in colonic epithelial cells surrounded by densely infiltrated leukocytes (Figure 6B). This finding suggests that the activation of Stat3 in inflamed colon was regulated by the activation of mTOR pathway. To determine the potential cross-talk between the mTOR and Stat3 pathways in inflamed colon, we examined the phosphorylation of Stat3 at Ser-727, an event that can be mediated by mTOR as well as by the mitogen-activated protein kinase pathway and is associated with the activation of the transcriptional regulatory activity of Stat3 toward its target genes.\textsuperscript{35,36} Nuclear staining of Stat3 phosphorylated at Ser-727 was observed in the colonic epithelial cells in the inflamed colon of Stat3-IKO mice (Figure 6D), in a pattern similar to that previously observed for the expression of pS6 and activated Stat3. Furthermore, treatment with Everolimus markedly reduced the phosphorylation of Stat3 at Ser-727 in these colonic epithelial cells (Figure 6C). These data indicate that the activation of Stat3 and its downstream factors in colonic epithelial cells during inflammation-induced tumorigenesis is regulated by cross-talk between mTOR and Stat3.

Abnormal Up-Regulation of mTOR-Stat3 Cross-Talk in the Colon of IBD Patients with Active Disease

To determine whether activation of the mTOR signaling pathway observed in Stat3-IKO mice mimics the inflammatory microenvironment in human IBD, the expression of pS6 in colon samples of four patients with active IBD as well as 40 human sporadic colorectal cancers was examined. IHC revealed positive staining of pS6 in the colonic epithelium in the inflamed colons from each of the four IBD patients, similar to that observed in Stat3-IKO mice.
The most intense staining was found in epithelial cells adjacent to areas of dense leukocytic infiltration (Figure 6E). In contrast, more than 80% of human sporadic colorectal cancer samples and normal colonic glands adjacent to the tumor were negative for pS6 (Supplemental Figure S4, a and b, see http://ajp.amjpathol.org). Six of 40 human sporadic tumor samples displayed a few clusters of pS6-positive cells in the tumor (Supplemental Figure S4, c and d, arrows, see http://ajp.amjpathol.org), and the density and distribution of these positive cells were similar to those observed in normal mouse colon (Figure 5, J and L). These data suggest that the activation of mTOR in colonic epithelial cells is specifically induced by inflammation in the colon of IBD patients. As we observed in Stat3-IKO colons, striking expression of Stat3 phosphorylated at Tyr-705 and Ser-727 was found in epithelial cells, often surrounded by prominent infiltration of leukocytes, in the inflamed colon of IBD patients (Figure 6, A and B). These data indicate that the both mTOR and Stat3 pathways, as well as their cross-talk signaling, are abnormally up-regulated as a result of inflammation in the inflamed colon of human IBD patients. Furthermore, our novel mouse model of inflammation-associated colorectal cancer closely mimics the abnormal signaling observed in colonic epithelium in IBD.

**Discussion**

In this study, we establish a novel mouse model of inflammation-associated colorectal cancer by inactivation of Stat3 specifically in hematopoietic cells. Similar to studies using IL10-deficient mice, this model exhibits the development of carcinoma as a consequence of inflammation initiated by immune dysregulation leading to microfloral intolerance, features that mimic human IBD. In contrast to the IL10 deficiency model, however, the new model permits us to observe tumor development in a genetically unmodified colonic epithelium, facilitating mechanistic studies of tumorigenesis. We have exploited this aspect of the model to demonstrate a role for mTOR-Stat3 signaling in inflammation-associated epithelial hyperproliferation and the disruption of epithelial homeostasis. Our findings with respect to mTOR and Stat3 activation in human IBD lend further support to the Stat3-IKO model as a mimic of human disease, as does our observation of a frequency of malignant progression in the Stat3-IKO mice similar to that in IBD patients.

Although the pathogenesis of IBD is largely unknown, dysregulation of mucosal immunity, abrogating the normal hyporeactive state with respect to the intestinal microflora, is believed to be the cause of IBD, a high-risk factor for tumor development. Inactivation of IL-10, a potent anti-inflammatory cytokine, has been closely linked to the development of IBD in humans. IL-10 acts as a potent inhibitor of the activation of macrophages, monocytes, and dendritic cells as well as the activation and function of natural killer cells and T and B lymphocytes. Stat3 is a major mediator of signaling downstream of IL-10, as well as that of multiple other cytokines and growth factors.

![Figure 6. mTOR-Stat3 cross-talk in human IBD and Stat3-IKO mouse. A-D: mTOR-Stat3 cross-talk in epithelial cells of Stat3-IKO colon. A and B: Immunohistochemical analysis for phosphorylated Stat3 (pStat3) (Tyr705) for colonic sections from a Stat3-IKO mouse treated with Everolimus (A) and an untreated Stat3-IKO littermate (B). C and D: Immunohistochemical analysis for pStat3 (Ser-727) in colonic sections from a Stat3-IKO mouse treated with Everolimus (C) and an untreated Stat3-IKO littermate (D). E-G: Colon sections from human IBD patients. Immunohistochemical analysis for pS6 (E), pStat3 (Tyr705) (F), and pStat3 (Ser-727) (G). Arrows pointed to leukocytic infiltration in the colon sections. Scale bar = 30 μm.](http://ajp.amjpathol.org)
Stat3-IKO mice overproduced proinflammatory cytokines, including tumor necrosis factor-α, IL-6, and IL-1α and β, when treated with the bacterial membrane component lipopolysaccharide (data not shown). Inflammatory macrophages are known to produce an array of cytokines and chemokines that promote infiltration and activation of other leukocytes, including lymphocytes, and that also promote angiogenesis, cell proliferation, and tissue damage.43,44 However, unlike IL-10−/− mice, in which malignant tumors develop in the inflamed intestine,9 no tumors were reported in mice with LysM- or Mx1-Cre-mediated Stat3 inactivation in myeloid cells. In contrast, we demonstrate the development of malignancy in Stat3-IKO mice, suggesting that Stat3 in both myeloid and lymphoid cells may serve as a control point for the development of malignant intestinal tumors. Because mice with T lymphocyte-specific depletion of Stat3 develop neither colitis nor colonic tumors,14 it is possible that cross-talk among stromal cells is needed for the promotion of tumorigenesis. Alternatively, differences between Csf1r and LysM promoter activation of the Cre recombinase in myeloid cells may account for the enhanced tumorigenesis in Stat3-IKO mice. The expression of LysM is chiefly in mature macrophages and neutrophils,45–47 whereas activation of the Csf1r promoter is initiated at an earlier stage of the myeloid lineage, although the activity in mature granulocytes is weak.16 Thus, the Csf1r promoter may direct Stat3 depletion to a different and perhaps broader range of myeloid cells compared with LysM, modulating the function of subpopulations of myeloid cells involved in tumor development. In addition, the difference between these promoters can be a matter of efficacy.

Although Csf1r-Cre led to a highly efficient inactivation of Stat3 in bone marrow-derived macrophages, we observed a high density of mononuclear cells with activated Stat3 among infiltrated stromal cells in the inflamed colon of Stat3-IKO mice, consistent with observations in human IBD colons that a large percentage of infiltrated CD68+ macrophages and CD4+ lymphocytes expressed activated Stat3.48–50 The subtypes of myeloid and lymphoid cells in which Stat3 is inactivated or, in contrast, in which Stat3 is activated in the inflamed colon in Stat3-IKO mice, has not yet been determined. We anticipate that resolution of this issue will identify a population of myeloid/lymphoid cells that plays a key role in regulating homoeostasis of mucosal immunity in the colon, with the consequence that disrupting the function of these cells initiates inflammation, and that different population(s) of leukocytes, with intact Stat3 that can respond to cytokines produced at the inflammatory site (eg, IL-6, IL-12, IL-10, vascular endothelial growth factor, and IL-23 that signal through Stat339,51), will respond to this cue that prompts their infiltration into the colon.

Consistent with findings in human IBD and IL-10-deficient mice,2 our study demonstrates that microflora plays a fundamental role in the development of inflammation in the colon. The response to antibiotic treatment and the increased density of bacteria in the stool of Stat3-IKO mice demonstrate quantitative and possible qualitative changes in the populations of microflora and reduced antibacterial ability of the mucosal immune system in this model. Recent studies have shown that the network of IL-23, IL-22, and IL-17 plays an important role in mucosal antimicrobial activities.52,53 On the stimulation of bacteria, dendritic cells can produce IL-22 directly or indirectly through activation of Th17 with IL-6 and IL-23 that promotes the antibacterial activities in the epithelial barrier.52,53 In the normal situation, when the inflammatory stimulus is removed, IL-10 and transforming growth factor-β are believed to play an immune-suppressive role to resolve the inflammation.51 Interestingly, most of the cytokines involved in this process, including IL-10, IL-6, IL-23, and IL-22, mediate their signals through Stat3.51–54. It is possible that inactivation of Stat3 in certain myeloid/lymphoid cells in Stat3-IKO mice perturbs this regulation and compromises the epithelial barrier, which may induce changes in the normal populations of microflora and even lead to expansion of pathogenic bacteria. In addition, the persistent inflammation, which is often accompanied by overproduction of various proteinases, reactive oxygen, and nitrogen, may physically damage the epithelial barrier in Stat3-IKO mice. The potential disruption of the IL-23/Th17 axis, which was shown to regulate restitution and repair in intestinal epithelium,51 by Stat3 inactivation may further enhance the damage of the epithelial barrier and cause the invasion of pathogenic bacteria. Therefore, it is likely that interruption of the intestinal epithelial barrier occurs in the colon of Stat3-IKO mice, which may promote bacterial translocation into mucosa and submucosa with subsequent infiltration of leukocytes and augmented inflammation.

A large percentage of Stat3-IKO mice developed hyperplastic and dysplastic lesions in the colon, with ~16% of the mice progressing to invasive carcinoma, a phenotype always associated with more extensive inflammation. Compared with other commonly used mouse models of inflammation-associated colorectal cancers, the frequency of carcinoma development in the Stat3-IKO mouse is relatively low, but it is close to the 18 to 20% tumor incidence in human IBD patients, which is similarly associated with prolonged and extensive colitis.55–57 In other widely used mouse models of chemically induced inflammation-associated colorectal cancer, nearly 100% of mice develop colon tumors.58–60 Although it is convenient for conducting investigations with these models, the inflammation is acute, resulting from mucosal damage, which may not recapitulate stochastic events involved in inflammation-associated cancers in the human.5 In contrast, tumors in Stat3-IKO mice are induced by spontaneous chronic inflammation, which may more closely model the dysregulation of mucosal immunity in the etiology of human IBD.5 The finding that treating Stat3-IKO mice with antibiotics reduced colonic inflammation, accompanied by a similar reduction in early stage tumor lesions, suggests that inflammation is a causal factor for the tumor development and persistence in the colon. This finding is consistent with previous reports that use of nonsteroidal anti-inflammatory drugs and treatment of experimental animal models with cyclooxygenase 2 or nuclear factor-kB inhibitors to inhibit inflammation lead to tumor regression.61–63 Our findings also further demon-
strate that tumor development in Stat3-IKO mice is caused by the abnormal mucosal immune response to microflora, as proposed for the pathogenesis of human IBD and associated colorectal cancers.2

By specifically targeting Stat3 depletion to the stromal compartment, the Stat3-IKO model, unlike IL-10-deficient models, allowed us to demonstrate that genetic disturbance of homeostasis in this compartment is sufficient for inflammation-associated tumorigenesis. Conversely, by preserving Stat3 function in epithelial cells, we were able to observe that epithelial Stat3, which is activated in human tumors associated with inflammation,64–66 was indeed activated in tumors of Stat3-IKO mice. Stat3 is persistently active in various human malignancies and possesses oncogenic potential,30 and the activation of Stat3 in human colorectal cancer cells is regulated by the microflora, as proposed for the pathogenesis of human IBD and associated colorectal cancers.2

The activation of Stat3 in human colorectal cancer cells is regulated by the microflora, as proposed for the pathogenesis of human IBD and associated colorectal cancers.2 However, a role for mTOR in inflammation-associated tumorigenesis. Conversely, by preserving Stat3 function in epithelial cells, we were able to observe that epithelial Stat3, which is activated in human tumors associated with inflammation,64–66 was indeed activated in tumors of Stat3-IKO mice. Stat3 is persistently active in various human malignancies and possesses oncogenic potential,30 and the activation of Stat3 in human colorectal cancer cells is regulated by the microflora, as proposed for the pathogenesis of human IBD and associated colorectal cancers.2

The factors that generate mTOR-Stat3 activation in epithelial cells in response to inflammation in Stat3-IKO mice have not been determined. A recent study in an irritant-induced colitis model provided evidence for IL-6 as a link between inflammation and Stat3 activation.74 The authors did not observe a dependence of mTOR activation on IL-6. It is possible that Stat3 activity represents the combined effect of IL-6 and a parallel pathway, yet to be determined, that regulates mTOR. Alternatively, because IL-6 is capable of up-regulating mTOR signaling86, the role of mTOR as an IL-6 effector may be dependent on the model studied. Future studies will be directed to the elucidation of factors governing the activation of these pathways in the inflamed colon. Although the etiology of IBD is not clear, interaction of the intestinal microflora with the gut immune system probably plays an important role.2,89,90 The data presented here suggest that the novel model established in this study mimics the modulation of the intestinal mucosal immune response by a complex interplay of genetic, microbial, and environmental factors in human IBD and points to mTOR as a potentially key mediator of these interactions.
Acknowledgments

We thank Dr. Rolf Sprengel (Max-Planck Institute for Medical Research) for the iCre fragment, Dr. Colin Stewart (NIH) for Stat3flox/mer mice, and Dr. Matthias M. Stadtfeld (Harvard Stem Cell Institute) for Rosa26-YFP mice. We thank Drs. Sandra Guilmart, Marta Flandez-Canet, Lidiya Klampfer, Anna Velcich, Ken Chen, Robert Odze, Kathryn E. Tanaka, and Larry Herbst for useful discussions; Michelle Houston, Elena Dhima, Mark Thomas, and James Lee for excellent technical support; the technical staffs of the Analytical Imaging Facility, Histotechnology and Comparative Pathology Facility, Transgenic Mouse Facility, and Fluorescence Activated Cell Sorter Facility of the Albert Einstein College of Medicine; and Mary Larocca and Kerry Victor for administrative support.

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