Can Scarring Be Turned Off?

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The waxing and waning of sequential events characterize the process of adult tissue repair: hemostasis, inflammation, fibrogenesis, angiogenesis, epithelization, and (in most tissues) scarring.1 Much of the analysis of regulation has focused on the activation of these events through local extrinsic signals impinging on transmembrane receptors. Indeed, these signals, whether they are cytokines, growth factors, arachidonic acid metabolites, or even simpler molecules such as nitric oxide, have been observed to undergo dynamic changes that are closely correlated with the activation of repair events, but questions arise. Are these repair processes attenuated merely by the cessation of signaling? Are there feedback mechanisms that govern healing outcomes through alternative pathways? A series of recent observations by Wells et al have begun to strengthen the notion that there are indeed pathways? A series of recent observations by Wells et al have begun to strengthen the notion that there are indeed "off" signals that operate during repair. Their current report2 on the effects of global CXCR3 deletion suggests that lack of this chemokine receptor results in protracted healing.

What Turns Healing Off?

Two decades of wound healing investigation have shown that the dynamic changes in wound organization are paralleled by changes in the abundance and/or expression of cytokines and growth factors. The discovery of these signal pathways, beginning with epidermal growth factor (EGF), has led to the development of diverse strategies to augment such signals where healing is impaired or to attenuate them where healing is excessive.3 This differential effect of the CXCR3 ligands was attributed to the inhibition of calpain proteinases in fibroblasts, thus preventing fibroblast detachment, and the activation of calpains in keratinocytes, thus promoting keratinocyte de-adhesion, presumably when IP-9 binding to CXCR3 drives Ca2+-dependent activation of μ-calpain (ie, calpain 1) via PLC-β3 signaling. This activation process would lead to cleavage of several components of the focal adhesion complex at the leading edge of the cell. Furthermore, EGF-induced motility is dependent on ERK-mediated activation of M-calpain (ie, calpain 2). In fibroblasts, however, IP-9 blocked EGF activation of M-calpain and conversely prevented cell detachment at the trailing edge.4

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These findings led the authors to propose that: (1) juxtacrine actions of epidermal IP-9 could promote epidermal resurfacing while paracrine action inhibited fibroblast and endothelial cell mobilization, and (2) concurrent endothelial cell expression of IP-10 could reinforce the inhibitory process in the dermal/mesenchymal compartment. Subsequently, knockdown of IP-9 expression in the epidermis of transgenic mouse via a keratin 5–driven antisense vector helped to parse out the relative effects of these two CXCR3 ligands. Epidermal healing was delayed in these mice, and the adjacent superficial dermis showed persistent hypercellularity and reduced organization as long as 60 days after injury. Despite the knockdown of the epidermal IP-9 expression, inflammation was not reduced. The interpretation of these ELR CXC cytokine (IP-9 and IP-10) knockout experiments could be confounded by the expression of these various ligands in different tissue compartments as well as the acute release of an additional CXCR3 ligand, PF4/CXCL4, after tissue injury.

In the global CXCR3 knockout mouse, wound healing is markedly impaired and delayed, with notable defects in the organization of the epidermis and its basement membrane up to 90 days after injury. The report in this issue describes the seeming reversal of the healing process when observation of wounds in these null mice continued out to 180 days. The authors observed epidermal thickening, poor organization of the dermal extracellular matrix, and concomitant defects in mechanical properties, which represent properties similar to human hypertrophic scars. The revival of an inflammatory response in the absence of infection shows that multiple aspects of the immune system were also affected by the loss of CXCR3. These findings suggest that the CXCR3 pathway, at least in mice, may be critical to involution and resolution of cutaneous healing.

There are certainly parallels and contrasts between the nature of the healing response in this model and the classical examples of excess cutaneous scarring in the human—hypertrophic scars and keloids. However, the persistence of incompletely reorganized wound sites and the regression of wound organization in this knockout model are quite remarkable.

It has been extremely difficult to find and use animal models to understand the cutaneous scarring process, although a wide variety of fibroses can be generated in cardiac, renal, pulmonary, or hepatic tissue. The only alternatives to date have been large excisions in the rabbit ear, the red Duroc pig, repeated bleomycin infiltration into mouse skin, and, most recently, progressive mechanical distension of incisional wounds in the mouse. It is possible that hypertrophy in these wounds is driven by a combination of an inappropriate mechanical environment and inflammatory mediators.

**CXCR3, IFN-γ, and Fibrosis**

The ELR CXC chemokines are induced by interferon (IFN)-γ, and they play roles in many aspects of inflammation and repair. The CXCR3 receptor is expressed on T-cells, NK cells, monocytes, dendritic cells, endothelial cells, fibroblasts, and keratinocytes. CXCR3 ligands act as potent leukocyte chemoattractants, and they are likely to modulate the abundance of NK and other T-cell populations at sites of acute and chronic injury. These chemokines are also known to inhibit angiogenesis through interaction with endothelial CXCR3. In the mouse model of bleomycin-induced pulmonary fibrosis, IP-10 administration reduced the severity of fibrosis, and knockout of CXCR3 led to a more severe fibrotic response and increased mortality without increased inflammatory cell recruitment. This report also proposed that early, self-amplifying induction of IFN-γ expression by CXCR3 signaling (via PF4/CXCL4) could be a critical step in limiting the fibrotic response. Recent analysis of an IP-10 knockout in a cardiac infarct model showed a delay in scar contraction, rapid thinning of the ventricular wall, and a shift in the composition of the infiltrating cell populations. These and many other studies indicate that signaling through the CXCR3 can influence many other cell types in the immune and inflammatory system. Thus, it is likely that global knockout of CXCR3 affects the tissue repair process at many levels and stages of repair. The delayed reappearance of a chronic wound phenotype in the present report may well reflect a contribution of the CXCR3 signal pathway in the immune system rather than resident epithelial and mesenchymal cells.

**Alternative Pathways**

The global nature of the CXCR3 knockout could certainly have an impact on cell types other than resident wound cells. The current work discusses potential effects on recruitment of endothelial and mesenchymal progenitors from the circulation, but the known participation of the CXCR3 circuitry in lymphocyte/T-cell and perhaps macrophage recruitment should not be overlooked. Both cell types are involved in the wound repair process. In the fibrotic lung, the immunomodulatory aspects of CXCR3 have been the predominant focus. The striking return of an inflammatory response 6 months after wounding is not typical of the hypertrophic scar. This phenomenon could well involve aberrant regulation of the immune system rather than further alteration of resident cell behavior.

Absent a cell-specific knockout of CXCR3, the strongest evidence for the importance of epidermal modulation of scar involution and remodeling comes from data obtained with epidermal-driven (keratin 5) overexpression of an IP-9 antisense construct. The current study provides data that partial suppression of IP-9 produces similar morphological effects, even at some distance below the epidermis. This finding does suggest that, at least in the mouse, CXCl1/CXCR3 interaction is the dominant mechanism for wound regression.

Hypertrophic scarring is a major complication of severe burn injuries, and burn surgeons have long been aware of the need for epithelial resurfacing to suppress the formation of excess connective tissue. The predominant modes of action of the CXCR3 system in formed elements appear to be cell motility and (endothelial) ap-
optosis. It is not certain from the present studies that these processes alone could account for the markedly reduced collagen content of wounds in the CXCR3-null mouse. A number of fibrotic mechanisms besides fibroblast motility have been proposed, including mechanical constraint, regulation of matrix metalloproteinases, and alteration of fibrogenic cytokine profiles. Other investigations of keratinocyte-fibroblast interactions have implicated other soluble mediators, including transforming growth factor (TGF)-β, fibroblast growth factor, interleukin (IL)-1, IL-7, and stratifin as agents that can modify the fibroblast phenotype. Many of these factors affect either the balance between synthesis and degradation of collagen or the rate of cell proliferation. It would be useful to undertake parallel studies, especially in clinical material, of the effects of the ELR CXCL-CXCR3 axis, these alternative signals, and the relative importance of their expression during normal and pathological wound healing.

**CXCR3 and Fibroplasia**

Previous studies in the CXCR3-null mouse certainly agree with the role of this receptor in limiting fibrosis. It has been exceedingly difficult to induce persistent hypertrophic scarring in animal skin by any means. The long-term response of the CXCR3-null wound does reflect several features of the hypertrophic scar, although it does not appear raised above the plane of the skin or hypervascularized. The CXCR3-null animals showed significantly increased, progressive, interstitial fibrosis without persistent inflammation in the pulmonary bleomycin model, whereas acute inflammation created patchy fibrosis in wild-type mice. These authors proposed that the suppression of an early burst of IFN-γ activity could have interrupted a positive feedback loop in which CXCL10/CXCR3 interpreted the protective effects of IP-10/CXCR3 signals to inhibit cell proliferation. It would be useful to undertake parallel studies, especially in clinical material, of the effects of the ELR CXCL-CXCR3 axis, these alternative signals, and the relative importance of their expression during normal and pathological wound healing.

**References**


